

Regulation of physiological responses to continuous and intermittent hypoxia by hypoxia-inducible factor 1

Gregg L. Semenza

Vascular Biology Program, Institute for Cell Engineering; Departments of Pediatrics, Medicine, Oncology and Radiation Oncology; and McKusick-Nathans Institute of Genetic Medicine, The Johns Hopkins University School of Medicine, Baltimore, MD, USA

Hypoxia-inducible factor 1 (HIF-1) is a heterodimeric transcription factor composed of HIF-1 α and HIF-1 β subunits that functions as a master regulator of oxygen homeostasis. Oxygen-dependent hydroxylation of HIF-1 α provides a mechanism that allows changes in oxygenation to be transduced to the nucleus, leading to changes in gene expression. Hypoxia-inducible factor 1 plays critical roles in development, physiology and disease pathogenesis. Analyses of mice that are heterozygous for a null allele at the locus encoding the HIF-1 α subunit have demonstrated that partial deficiency of HIF-1 is sufficient to impair multiple physiological responses to continuous and intermittent hypoxia.

(Received 17 April 2006; accepted after revision 10 May 2006; first published online 1 June 2006)

Corresponding author G. L. Semenza: Broadway Research Building, Suite 671, 733 N. Broadway, Baltimore, MD 21205, USA. Email: gsemenza@jhmi.edu

Oxygen homeostasis

Humans and other vertebrates have evolved complex circulatory and respiratory systems to insure the delivery of adequate supplies of O₂ to meet metabolic demands of every cell in the organism. Principal among these demands is the requirement for O₂ as ultimate electron acceptor in the process of oxidative phosphorylation, which generates sufficient ATP from the catabolism of glucose and fatty acids to maintain the complex nature of vertebrate structure and function. However, the need for O₂ as metabolic substrate is opposed by the inherent risk of oxidative damage to cellular macromolecules. Because of the dual character of O₂ as an essential but potentially toxic molecule, its concentration within cells of a healthy organism is maintained within a narrow range that optimally balances supply and demand.

Identification of hypoxia-inducible factor 1 (HIF-1)

At the molecular level, oxygen homeostasis is achieved by acute adaptive responses, which involve changes in the activity of pre-existing proteins, and chronic responses, which involve changes in gene expression leading to new protein synthesis. The transcriptional factor HIF-1 is a master regulator of oxygen homeostasis, HIF-1 was discovered through studies analysing the molecular mechanisms regulating expression of the human

gene encoding erythropoietin (EPO), the glycoprotein hormone that controls red blood cell production and, thus, blood O₂-carrying capacity.

These studies identified a *cis*-acting hypoxia-response element (HRE) in the 3'-flanking region of the *EPO* gene (Beck *et al.* 1991; Semenza *et al.* 1991). Similar results were reported for the mouse *Epo* gene (Pugh *et al.* 1991). When this HRE was inserted into a heterologous reporter gene and transfected into human cells, exposure of the cells to hypoxic conditions (1% O₂) resulted in a dramatic increase in transcription of the reporter gene, similar to that of the endogenous *EPO* gene. Deletion analysis localized the minimal HRE to a 33-base-pair (bp) sequence, and mutagenesis identified a sequence, 5'-CTACGTGCT-3', that when mutated resulted in loss of HRE activity (Semenza & Wang, 1992). A nuclear factor designated HIF-1 was identified, which was present in hypoxic cells but not in cells incubated under standard tissue culture conditions of 95% air and 5% CO₂ (20% O₂) and which bound to a double-stranded oligonucleotide containing the wild-type sequence but not to an oligonucleotide containing a 3 bp mutation in the DNA sequence (Semenza & Wang, 1992).

The differential binding of HIF-1 to the wild-type and mutant sequences was used as a positive and negative selection, respectively, to perform a biochemical purification of HIF-1 by DNA affinity chromatography from 100 l of human HeLa cells grown in suspension

culture (Wang & Semenza, 1995), leading to the isolation of nucleic acid sequences encoding the HIF-1 α and HIF-1 β subunits (Wang *et al.* 1995) and the generation of monoclonal antibodies that recognize the cognate proteins (Zhong *et al.* 1999; Talks *et al.* 2000; Zagzag *et al.* 2000). The utilization of these tools has led to a revolution in the molecular analysis of O₂ homeostasis in health and disease.

Transducing changes in oxygenation to changes in gene expression

Analysis of nuclear extracts prepared from HeLa cells exposed to precisely defined O₂ concentrations revealed that reduction from 20 to 6% O₂ resulted in a twofold increase in HIF-1 α protein levels, whereas below 6% O₂, HIF-1 α levels increased exponentially, with a half-maximal response at 1.5–2% O₂ and a maximal response at 0.5% O₂, with HIF-1 α protein levels that were ~10-fold greater than those in cells at 6% O₂ (Jiang *et al.* 1996). The levels of HIF-1 β in nuclear extracts of hypoxic cells increased only modestly, an effect that was later shown to result from an increased loss of the protein during preparation of nuclear extracts from non-hypoxic cells rather than an actual increase in protein levels within hypoxic cells (Chilov *et al.* 1999).

Regulation of HIF-1 α protein levels is effected by O₂-dependent ubiquitination and proteasomal degradation (Salceda & Caro, 1997; Huang *et al.* 1998; Kallio *et al.* 1999), which is mediated by the binding of the von Hippel-Lindau (VHL) tumour suppressor protein (Maxwell *et al.* 1999). VHL binding to HIF-1 α is triggered by the hydroxylation of two proline residues (Pro402 and Pro564 in human HIF-1 α ; Epstein *et al.* 2001; Ivan *et al.* 2001; Jaakkola *et al.* 2001; Yu *et al.* 2001). The HIF-1 α prolyl hydroxylases that perform this modification (Bruick & McKnight, 2001; Epstein *et al.* 2001) utilize molecular O₂ and α -ketoglutarate as substrates and generate succinate as a byproduct (Hewitson *et al.* 2004). Under conditions of severe hypoxia, substrate (O₂) deprivation is sufficient to inhibit the hydroxylation reaction, whereas under conditions of moderate hypoxia, the mitochondrial generation of reactive O₂ species is also required to inhibit the hydroxylases (Brunelle *et al.* 2005; Guzy *et al.* 2005; Mansfield *et al.* 2005). The O₂-dependent hydroxylation of HIF-1 α provides a molecular mechanism by which changes in oxygenation can be transduced to the nucleus via HIF-1, leading to changes in gene expression.

HIF-1 is required for embryonic development

Homologous recombination in mouse embryonic stem cells was used to generate a null allele at the *Hif1a* locus encoding HIF-1 α (Iyer *et al.* 1998). Mouse

embryos homozygous for the null allele arrested in their development by embryonic day 8.5 and died by day 10.5 with cardiac malformations, vascular regression and mesenchymal cell death (Iyer *et al.* 1998; Kotch *et al.* 1999). Embryonic lethality of HIF-1 α -null embryos was subsequently confirmed by independent mouse lines generated in two other laboratories (Carmeliet *et al.* 1998; Ryan *et al.* 1998).

Involvement of HIF-1 in hypoxia-induced pulmonary vascular remodelling

The embryonic lethality of HIF-1 α -null mice precluded analysis of the effects of complete HIF-1 α deficiency on postnatal physiology. However, the heterozygous-null mice, which develop normally and are indistinguishable from wild-type littermates, have impaired physiological responses to hypoxia and ischaemia. Exposure of wild-type littermates to continuous hypoxia (10% O₂ for 3 weeks) results in the development of pulmonary hypertension, which is manifested by right ventricular hypertrophy, elevated pulmonary artery pressure and increased medial wall thickness of pulmonary arterioles. All of these responses were impaired in heterozygotes (Yu *et al.* 1999). Electrophysiological analysis of pulmonary artery smooth muscle cells (PASMC) isolated from normoxic and hypoxic wild-type and heterozygous-null (HET) mice revealed that the hypoxia-induced hypertrophy and depolarization of these cells was impaired in HET mice (Shimoda *et al.* 2001). The depolarization of PASMCs from hypoxic wild-type mice was associated with a reduction in voltage-gated K⁺ channel currents, an effect that was not observed in PASMCs from hypoxic HET mice (Shimoda *et al.* 2001). Depolarization of PASMCs from hypoxic wild-type mice was associated with increased intracellular Ca²⁺ concentrations, an effect that was lost in HET mice. Taken together, these data indicate that HIF-1 plays a critical role in the pathophysiology of hypoxia-induced pulmonary hypertension.

HIF-1 is required for ventilatory responses to hypoxia mediated by the carotid body

The carotid bodies sense arterial partial pressure of O₂ and respond to hypoxaemia by activating neural pathways that stimulate increases in blood pressure, heart rate and ventilation. Ventilatory adaptation to chronic hypoxia, in which exposure to ambient hypoxia for several days results in an augmented ventilatory response to a subsequent acute hypoxic stimulus, is believed to be mediated by the carotid bodies. While HET mice manifested ventilatory responses to an acute hypoxic challenge, the ventilatory adaptation to chronic hypoxia was lost (Kline *et al.* 2002), suggesting a loss of carotid body function.

Vagotomy, which interrupts neurotransmission from oxygen-sensing cells in the aorta (aortic bodies) but not from the carotid body, did not affect the augmentation of phrenic nerve activity in response to an acute hypoxic stimulus in wild-type mice, whereas in HET mice, phrenic nerve activity in response to hypoxia was lost, suggesting that acute O₂ sensing in HET mice was principally mediated by aortic bodies, whereas the carotid bodies performed this function in wild-type mice.

Single fibre recording of sinus nerve activity from carotid bodies isolated from HET mice revealed a complete loss of neural activity in response to hypoxia. In contrast, sinus nerve activity stimulated by cyanide application was identical in wild-type and HET mice, as was carotid body histology, including glomus cell morphometry. Thus, partial HIF-1 α deficiency results in a specific and virtually complete loss of carotid body oxygen sensing and/or signal transduction. In HET mice, aortic bodies compensate for the lack of carotid body function, similar to the recovery of oxygen sensing that is observed after carotid denervation. Thus, the carotid bodies are particularly dependent upon normal levels of HIF-1 α expression for their physiological function as arterial oxygen sensors.

HIF-1 is required for cardiovascular and ventilatory responses to intermittent hypoxia

Individuals with obstructive sleep apnoea are subjected to chronic intermittent hypoxia during sleep. Exposure of mice to chronic intermittent hypoxia induces ventilatory adaptation to acute hypoxia in wild-type but not in HET mice, whereas the ventilatory adaptation to acute hypercapnia that is induced by chronic intermittent hypoxia is present in both wild-type and HET mice (Peng YJ, Yuan G, Ramakrishnan D, Sharma SD, Bosch-Marcé M, Kumar GK, Semenza GL & Prabhakar NR, unpublished observations).

In contrast to chronic continuous hypoxia, which results in the development of pulmonary hypertension, chronic intermittent hypoxia results in the development of systemic hypertension. Exposure of mice to chronic intermittent hypoxia for 10 days induces elevated serum noradrenaline levels and systemic hypertension in wild-type mice but not in HET mice (Peng YJ, Yuan G, Ramakrishnan D, Sharma SD, Bosch-Marcé M, Kumar GK, Semenza GL & Prabhakar NR, unpublished observations).

In addition to the development of systemic hypertension, individuals with intermittent hypoxia owing to sleep apnoea develop a metabolic syndrome consisting of insulin resistance, hypercholesterolaemia and hypertriglyceridaemia. These responses are also observed in wild-type mice exposed to intermittent hypoxia for 5 days, whereas in HET mice impaired development of hypertriglyceridaemia is associated with

impaired activation of sterol response element binding protein 1, a key activator of triglyceride synthesis (Li *et al.* 2006).

The studies described above have demonstrated that partial deficiency of HIF-1 α results in impaired physiological responses to chronic exposure to both continuous and intermittent hypoxia. Major causes of continuous and intermittent hypoxia are chronic obstructive lung disease resulting from smoking and obstructive sleep apnoea associated with obesity, respectively, which have arisen only recently in the history of *Homo sapiens* and thus have not been subjected to extensive natural selection. As a result, the physiological systems that have evolved to mediate adaptive responses and protect the organism against causes of hypoxia that would have been commonly encountered during most of human history, such as traumatic blood loss and acute pneumonia, mediate maladaptive responses to these novel stimuli. Further studies are needed to determine whether these maladaptive responses can be inhibited, e.g. by pharmacological inhibition of HIF-1 (Semenza, 2006), without interfering with other critical aspects of oxygen homeostasis that are also controlled by HIF-1.

References

- Beck I, Ramirez S, Weinmann R & Caro J (1991). Enhancer element at the 3'-flanking region controls transcriptional response to hypoxia in the human erythropoietin gene. *J Biol Chem* **266**, 15563–15566.
- Bruick RK & McKnight SL (2001). A conserved family of prolyl-4-hydroxylases that modify HIF. *Science* **294**, 1337–1340.
- Brunelle JK, Bell EL, Quesada NM, Vercauteren K, Tiranti V, Zeviani M *et al.* (2005). Oxygen sensing requires mitochondrial ROS but not oxidative phosphorylation. *Cell Metab* **1**, 409–414.
- Carmeliet P, Dor Y, Herbert JM, Fukumura D, Brusselmans K, Dewerchin M *et al.* (1998). Role of HIF-1 α in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature* **394**, 485–490.
- Chilov D, Camenisch G, Kvietikova I, Ziegler U, Gassmann M & Wenger RH (1999). Induction and nuclear translocation of hypoxia-inducible factor-1 (HIF-1): heterodimerization with ARNT is not necessary for nuclear accumulation of HIF-1 α . *J Cell Sci* **112**, 1203–1212.
- Epstein AC, Gleadle JM, McNeill LA, Hewitson KS, O'Rourke J, Mole DR *et al.* (2001). *C. elegans* EGL-9 and mammalian homologs define a family of dioxygenases that regulate HIF by prolyl hydroxylation. *Cell* **107**, 43–54.
- Guzy RD, Hoyos B, Robin E, Chen H, Liu L, Mansfield KD *et al.* (2005). Mitochondrial complex III is required for hypoxia-induced ROS production and cellular oxygen sensing. *Cell Metab* **1**, 401–408.
- Hewitson KS, McNeill LA & Schofield CJ (2004). Modulating the hypoxia-inducible factor signaling pathway. *Curr Pharm Des* **10**, 821–833.

- Huang LE, Gu J, Schau M & Bunn HF (1998). Regulation of hypoxia-inducible factor 1 α is mediated by an O₂-dependent degradation domain via the ubiquitin-proteasome pathway. *Proc Natl Acad Sci U S A* **95**, 7987–7992.
- Ivan M, Kondo K, Yang H, Kim W, Valiando J, Ohh M *et al.* (2001). HIF α targeted for VHL-mediated destruction by proline hydroxylation: implications for O₂ sensing. *Science* **292**, 464–468.
- Iyer NV, Kotch LE, Agani F, Leung SW, Laughner E, Wenger RH *et al.* (1998). Cellular and developmental control of O₂ homeostasis by hypoxia-inducible factor 1 α . *Genes Dev* **12**, 149–162.
- Jaakkola P, Mole DR, Tian YM, Wilson MI, Gielbert J, Gaskell SJ *et al.* (2001). Targeting of HIF α to the von Hippel-Lindau ubiquitylation complex by O₂-regulated prolyl hydroxylation. *Science* **292**, 468–472.
- Jiang BH, Semenza GL, Bauer C & Marti HH (1996). Hypoxia-inducible factor 1 levels vary exponentially over a physiologically relevant range of O₂ tension. *Am J Physiol* **271**, C1172–C1180.
- Kallio PJ, Wilson WJ, O'Brien S, Makino Y & Poellinger L (1999). Regulation of the hypoxia-inducible transcription factor 1 α by the ubiquitin-proteasome pathway. *J Biol Chem* **274**, 6519–6525.
- Kline DD, Peng Y, Manalo DJ, Semenza GL & Prabhakar NR (2002). Defective carotid body function and impaired ventilatory responses to chronic hypoxia in mice partially deficient for hypoxia-inducible factor 1 α . *Proc Natl Acad Sci U S A* **99**, 821–826.
- Kotch LE, Iyer NV, Laughner E & Semenza GL (1999). Defective vascularization of HIF-1 α -null embryos is not associated with VEGF deficiency but with mesenchymal cell death. *Dev Biol* **209**, 254–267.
- Li J, Bosch-Marce M, Nanayakkara A, Savransky V, Fried SK *et al.* (2006). Altered metabolic responses to intermittent hypoxia in mice with partial deficiency of hypoxia-inducible factor 1 α . *Physiol Genomics* **25**, 450–457.
- Mansfield KD, Guzy RD, Pan Y, Young RM, Cash TP *et al.* (2005). Mitochondrial dysfunction resulting from loss of cytochrome c impairs cellular oxygen sensing and hypoxic HIF- α activation. *Cell Metab* **1**, 393–399.
- Maxwell PH, Wiesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME *et al.* (1999). The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature* **399**, 271–275.
- Pugh CW, Tan CC, Jones RW & Ratcliffe PJ (1991). Functional analysis of an oxygen-regulated transcriptional enhancer lying 3' to the mouse erythropoietin gene. *Proc Natl Acad Sci U S A* **88**, 10553–10557.
- Ryan HE, Lo J & Johnson RS (1998). HIF-1 α is required for solid tumor formation and embryonic vascularization. *EMBO J* **17**, 3005–3015.
- Salceda S & Caro J (1997). Hypoxia-inducible factor 1 α (HIF-1 α) protein is rapidly degraded by the ubiquitin-proteasome system under normoxic conditions. Its stabilization by hypoxia depends on redox-induced changes. *J Biol Chem* **272**, 22642–22647.
- Semenza GL (2006). Development of novel therapeutic strategies that target HIF-1. *Expert Opin Ther Targets* **10**, 267–280.
- Semenza GL, Neifelt MK, Chi SM & Antonarakis SE (1991). Hypoxia-inducible nuclear factors bind to an enhancer located 3' to the human erythropoietin gene. *Proc Natl Acad Sci U S A* **88**, 5680–5684.
- Semenza GL & Wang GL (1992). A nuclear factor induced by hypoxia via de novo protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. *Mol Cell Biol* **12**, 5447–5454.
- Shimoda LA, Manalo DJ, Sham JSK, Semenza GL & Sylvester JT (2001). Partial HIF-1 α deficiency impairs pulmonary arterial myocyte electrophysiological responses to chronic hypoxia. *Am J Physiol* **281**, L202–L208.
- Talks KL, Turley H, Gatter KC, Maxwell PH, Pugh CW *et al.* (2000). The expression and distribution of the hypoxia-inducible factors HIF-1 α and HIF-2 α in normal human tissues, cancers, and tumor-associated macrophages. *Am J Pathol* **157**, 411–421.
- Wang GL, Jiang BH, Rue EA & Semenza GL (1995). Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O₂ tension. *Proc Natl Acad Sci U S A* **92**, 5510–5514.
- Wang GL & Semenza GL (1995). Purification and characterization of hypoxia-inducible factor 1. *J Biol Chem* **270**, 1230–1237.
- Yu AY, Shimoda LA, Iyer NV, Huso DL, Sun X, McWilliams R, Beaty T *et al.* (1999). Impaired physiological responses to chronic hypoxia in mice partially deficient for hypoxia-inducible factor 1 α . *J Clin Invest* **103**, 691–696.
- Yu F, White SB, Zhao Q & Lee FS (2001). HIF-1 α binding to VHL is regulated by stimulus-sensitive proline hydroxylation. *Proc Natl Acad Sci U S A* **98**, 9630–9635.
- Zagzag D, Zhong H, Scalzitti JM, Laughner E, Simons JW & Semenza GL (2000). Expression of hypoxia-inducible factor 1 α in human brain tumors: association with angiogenesis, invasion, and progression. *Cancer* **88**, 2606–2618.
- Zhong H, De Marzo AM, Laughner E, Lim M, Hilton DA, Zagzag D *et al.* (1999). Overexpression of hypoxia-inducible factor 1 α in common human cancers and their metastases. *Cancer Res* **59**, 5830–5835.

Acknowledgements

I thank the many members of my laboratory who contributed to the work described in this review. The animal physiology studies would not have been possible without the privilege of collaborating with Nanduri Prabhakar of Case Western Reserve University, Larissa Shimoda and Seva Polotsky of Johns Hopkins University, and the talented members of their laboratories.